mations with respect to the $\mathrm{C}(1)-\mathrm{C}(2)$ and $\mathrm{C}(8)$ $\mathrm{C}(9)$ bonds respectively. These conformations bring the $\mathrm{N}(1)-\mathrm{H}$ proton and the $\mathrm{N}(2)$ lone pair into a favourable position to form an intramolecular hydrogen bond. However, in DMAN these conformations are such that at each N atom one methyl group is almost eclipsed with either $\mathrm{C}(1)-\mathrm{C}(2)$ or $\mathrm{C}(8)-\mathrm{C}(9)$ (torsion angles -18 and $-19 \cdot 4^{\circ}$ respectively) while the other methyl groups have torsion angles of 120 and $119^{\circ}$ respectively.

## References

Alder, R. W., Bowman, P. S., Steele, W. R. S. \& Wintermann, D. R. (1968). Chem. Commun. pp. 723-724.

Alder, R. W., Goode, N. C., Miller, N., Hibbert, F., Hunte, K. P. P. \& Robbins, H. J. (1978). Chem. Commun. pp. 89-90.
allen, F. H., Kennard, O. \& Taylor, R. (1983). Acc. Chem. Res. 16, 146-153.
Awwal, A. \& Hibbert, F. (1977). J. Chem. Soc. Perkin Trans. 2, pp. 152-156.
Brown, D. A., Clegg, W., Colquhoun, H. M., Daniels, J. A., Stephenson, I. R. \& Wade, K. (1987). Chem. Commun. pp. 889-891.
Chonacki, H., Laskowski, Z. \& Malarski, Z. (1983). J. Mol. Struct. 102, 251-255.

Cromer, D. T. \& Liberman, D. (1970). J. Chem. Phys. 53 1891-1898.
Cromer, D. T. \& Mann, J. B. (1968). Acta Cryst. A24, 321-324.
Einspahr, H., Robert, J. B., Marsh, R. E. \& Roberts, J. D. (1973). Acta Cryst. B29, 1611-1617.

Glowiak, T., Malarski, Z., Sobczyk, L. \& Grech, E. (1987). J. Mol. Struct. 157, 329-337.
Grech, E., Malarski, Z. \& Sobczyk, L. (1985). J. Mol. Struct. 129, 35-43.
Groot, R. L. de \& Sikkema, D. J. (1976). Recl Trav. Chim. Pays-Bas, 95, 10-14.
Hamann, S. D. \& Linton, M. (1976). Austr. J. Chem. 29, 1641-1647.
Hibbert, F. (1974). J. Chem. Soc. Perkin Trans. 2, 1862-1866.
Miller, P. K., Abney, K. D., Rappé, A. K., Anderson, O. P. \& Strauss, S. H. (1988). Inorg. Chem. 27, 2255-2261.
Pawley, G. S. \& Yeats, E. A. (1969). Acta Cryst. B25, 20092013.

Pyzalska, D., Pyzalski, R. \& Borowiak, T. (1983). J. Cryst. Spectrosc. Res. 13, 211-220.
Sheldrick, G. M. (1976). SHELX76. Program for crystal structure determination. Univ. of Cambridge, England.
Sheldrick, G. M. (1986). SHELXS86. Program for the solution of crystal structures. Univ. of Göttingen, Germany.
Spek. A. L. (1982). The EUCLID Package. In Computational Crystallography, edited by D. Sayre, p. 528. Oxford: Clarendon Press.
Truter, M. R. \& Vickery, B. L. (1972). J. Chem. Soc. Dalton Trans. pp. 395-403.

Acta Cryst. (1991). C47, 810-812

# Structure of Lantadene A, the Major Triterpenoid of Lantana Camara, Red Variety* 

By Vasantha Pattabhi $\dagger$ and N. Sukumar<br>Department of Crystallography and Biophysics, University of Madras, Madras-600 025, India

and O. P. Sharma<br>Biochemistry Laboratory, Indian Veterinary Research Institute, Regional Station, Palampur, Kanagara Valley, Himachala Pradesh-176 061, India

(Received 5 June 1990; accepted 2 August 1990)


#### Abstract

Lantadene A, [22 $\beta(Z)]$-2-methylisocro-tonoyloxy-3-oxoolean-12-en-28-oic acid, is the major triterpenoid constituent of L. Camara, red variety. Mixed toxin preparation from lantana leaves has been found to exist in two molecular forms, of which only one was found to be hepatotoxic to guinea pigs. The structure of form ( I ), which is not hepatotoxic to guinea pigs, is reported. $\mathrm{C}_{35} \mathrm{H}_{52} \mathrm{O}_{5}, M_{r}=552.35$, orthorhombic, $\quad P 2_{1} 2_{1} 2_{1}, \quad a=12.494$ (2),$\quad b=$ 15.790 (2),$c=16.367$ (2) $\AA, \quad V=3229 \cdot 0 \AA^{3}, \quad Z=4$,


* DCB Contribution No. 757.
$\dagger$ To whom correspondence should be addressed.
$D_{x}=1.14 \mathrm{~g} \mathrm{~cm}^{-3}, \quad D_{m}=1.15 \mathrm{~g} \mathrm{~cm}^{-3}, \quad \lambda(\mathrm{Cu} \mathrm{K} \mathrm{\alpha})=$ $1.5418 \AA, \quad \mu=5.9 \mathrm{~cm}^{-1}, \quad F(000)=1208, \quad T=$ $296(1) \mathrm{K}, \quad R=0.061$ for 3152 unique observed reflections. The $A / B$ and $B / C$ rings are trans fused while the $D / E$ rings are cis fused. The packing of the molecule is stabilized by $\mathrm{O}-\mathrm{H} \cdots \mathrm{O}$ hydrogen bonds.

Introduction. Ingestion of lantana foliage causes cholestasis and hepatotoxicity in animals (Sharma, Makkar \& Dawra, 1988). Consensus did not exist regarding the capacity of lantadene A, which is the major constituent of L. Camara, red variety (Sharma, Dawra \& Makkar, 1987), to elicit hepato-
toxicity (Sharma, Makkar \& Dawra, 1988). Crystal polymorphism and difference in molecular forms of drugs and xenobiotics are known to influence their interactions with biomolecules and receptors (Ravin, 1985). Recently, we reported a mixed toxin preparation from lantana leaves which existed in two molecular forms and only one of them was found to be hepatotoxic to guinea pigs (Sharma, Dawra \& Makkar, 1988). The present study was undertaken to investigate the X-ray structure of lantadene A [form (I)] for its further use in understanding pharmacodynamics and the spatial aspects of its molecular interactions in the vicinity of biomolecules and biomembranes.


Experimental. Crystals of lantadene A were obtained from a mixture of $80 \%$ benzene, $10 \%$ acetone and $10 \%$ 2-propanol. Intensity data were collected on a colorless parallelepiped crystal, $0.35 \times 0.35 \times$ 0.30 mm , on an Enraf-Nonius CAD-4 diffractometer in the $\omega / 2 \theta$ scan mode in the range $2 \leq 2 \theta \leq$ $140^{\circ}, \quad 0 \leq h \leq 15, \quad 0 \leq k \leq 19, \quad 0 \leq l \leq 19 \quad$ with graphite-monochromated $\mathrm{Cu} K \alpha$ radiation. Data were corrected for direct-beam polarization and Lorentz effects. Empirical absorption correction with maximum and minimum correction factors 0.999 and 0.982 , respectively, was applied. Out of 3492 reflections measured, 3426 were unique and 3152 observed with $I \geq 2 \cdot 5 \sigma(I)$. Cell constants were obtained from 20 reflections in the range $14<\theta<38^{\circ}$. Three standard reflections monitored every 2 h did not show any variation in intensity. Aperture width $=(4+$ $2 \tan \theta) \mathrm{mm}$, maximum time spent on any reflection was 60 s and the background count was half the scan time.

Structure solution was by direct methods, 22 out of 52 H atoms were located from $\Delta \rho$ maps and the rest (barring the hydroxyl H atom) were geometrically fixed and refined isotropically only for two cycles. The non- H atoms were refined anisotropically on $F_{o}$ to a final $R=0.061$ and $w R=0.090$, with individual weighting scheme based on counting statistics, where $w=4 F_{o}{ }^{2} / \sigma^{2}\left(F_{o}^{2}\right), \quad \sigma\left(F_{o}^{2}\right)=\left[\sigma^{2}(I)+\right.$ $\left(0.05 I^{2}\right]^{1 / 2} / \mathrm{Lp} . \quad(\Delta / \sigma)_{\max }=0.04, \quad S=2.77$ for 361 parameters. Final map had $(\Delta \rho)_{\max } /(\Delta \rho)_{\min }=0.34 /$

Table 1. Positional parameters and $B_{\mathrm{eq}}$ values of non- H atoms with e.s.d.'s in parentheses

Anisotropically refined atoms are given in the form of the isotropic equivalent displacement parameter defined as: $(4 / 3)\left[a^{2} B(1,1)+b^{2} B(2,2)+c^{2} B(3,3)\right.$ $+a b(\cos \gamma) B(1,2)+a c(\cos \beta) B(1,3)+b c(\cos \alpha) B(2,3)]$.

|  | $x$ | $y$ | $z$ | $B_{\text {eq }}\left(\AA^{2}\right)$ |
| :---: | :---: | :---: | :---: | :---: |
| O1 | 0.2224 (2) | 0.0196 (1) | 0.7969 (1) | 3.44 (4) |
| O 2 | $0 \cdot 2918$ (3) | 0.7830 (1) | 1.0433 (2) | $5 \cdot 20$ (6) |
| O3 | 0.3079 (2) | 0.1564 (2) | 0.6778 (2) | $4 \cdot 28$ (5) |
| 04 | $0 \cdot 1781$ (2) | 0.2136 (2) | 0.7524 (2) | 4.43 (5) |
| O5 | 0.2823 (3) | -0.0968 (2) | 0.7350 (2) | $7 \cdot 15$ (8) |
| Cl | $0 \cdot 2528$ (3) | 0.5590 (2) | 1.0601 (2) | 4.79 (7) |
| C2 | 0.2731 (5) | 0.6466 (2) | 1.0930 (2) | 6.5 (1) |
| C3 | 0.3189 (3) | 0.7090 (2) | 1.0374 (2) | 3.89 (6) |
| C4 | 0.4009 (3) | 0.6833 (2) | 0.9738 (2) | 3.50 (6) |
| C5 | 0.3956 (3) | 0.5854 (2) | 0.9570 (2) | 2.75 (5) |
| C6 | 0.4426 (3) | $0 \cdot 5610$ (2) | 0.8742 (2) | 3.35 (6) |
| C7 | 0.4594 (3) | 0.4666 (2) | 0.8686 (2) | 3.17 (6) |
| C8 | 0.3556 (2) | 0.4151 (2) | 0.8802 (2) | 2.45 (5) |
| C9 | 0.2945 (3) | 0.4465 (2) | 0.9568 (2) | 2.92 (6) |
| C10 | 0.2842 (3) | 0.5453 (2) | 0.9703 (2) | 3.00 (6) |
| C11 | 0.1850 (3) | 0.4019 (2) | 0.9608 (3) | 4.91 (8) |
| C12 | 0.1947 (3) | $0 \cdot 3088$ (2) | 0.9435 (3) | $4 \cdot 26$ (7) |
| C 13 | 0.2809 (3) | $0 \cdot 2690$ (2) | 0.9145 (2) | $2 \cdot 82$ (5) |
| C 14 | 0.3837 (2) | 0.3173 (2) | 0.8944 (2) | $2 \cdot 51$ (5) |
| C15 | 0.4367 (3) | 0.2799 (2) | 0.8167 (2) | $3 \cdot 12$ (6) |
| C16 | 0.4400 (3) | 0.1822 (2) | 0.8162 (2) | $2 \cdot 82$ (5) |
| C17 | 0.3262 (2) | $0 \cdot 1446$ (2) | 0.8212 (2) | 2.50 (5) |
| C18 | 0.2731 (3) | 0.1729 (2) | 0.9025 (2) | 2.75 (5) |
| C 19 | $0 \cdot 3192$ (3) | 0.1250 (2) | 0.9766 (2) | $3 \cdot 39$ (6) |
| C20 | $0 \cdot 3185$ (3) | 0.0278 (2) | 0.9701 (2) | $3 \cdot 22$ (6) |
| C21 | 0.3737 (3) | 0.0027 (2) | 0.8900 (2) | $3 \cdot 10$ (6) |
| C22 | 0.3317 (3) | 0.0470 (2) | 0.8137 (2) | 2.89 (6) |
| C23 | 0.3898 (4) | 0.7392 (2) | 0.8987 (2) | 5.66 (9) |
| C24 | 0.5108 (4) | 0.7045 (3) | 1.0148 (4) | 7.0 (1) |
| C25 | 0.1978 (3) | 0.5877 (2) | 0.9180 (3) | 4.57 (8) |
| C26 | 0.2874 (3) | 0.4244 (2) | 0.8018 (2) | 3.95 (7) |
| C27 | 0.4644 (3) | $0 \cdot 3054$ (2) | 0.9649 (2) | 3.66 (7) |
| C28 | 0.2610 (3) | $0 \cdot 1756$ (2) | 0.7482 (2) | $3 \cdot 28$ (6) |
| C29 | 0.2044 (3) | -0.0087 (2) | 0.9772 (3) | $4 \cdot 10$ (7) |
| C30 | 0.3846 (3) | -0.0097 (2) | 1.0414 (2) | 4.66 (7) |
| C31 | $0 \cdot 2086$ (3) | -0.0551 (2) | 0.7599 (2) | $4 \cdot 46$ (7) |
| C32 | 0.0908 (4) | -0.0745 (3) | 0.7528 (3) | 6.38 (9) |
| C33 | 0.0593 (5) | -0.1523 (3) | 0.7311 (4) | $9 \cdot 2$ (1) |
| C34 | 0.1181 (9) | -0.2272 (4) | 0.7202 (6) | 14.2 (3) |
| C35 | 0.0144 (4) | -0.0010 (4) | 0.7666 (4) | $8 \cdot 5$ (1) |

$-0.3 \mathrm{e} \AA^{-3}$. Atomic scattering factors were from International Tables for X-ray Crystallography (1974, Vol. IV, pp. 99, 149). All calculations were carried out on a VAX 11/730 computing system using the SDP package (Frenz, 1978). The final atomic coordinates are listed in Table 1.*

Discussion. Bond lengths and angles (Table 2) agree with the values observed in similar structures such as maytenfolic acid, maytenfoliol (Nozaki, Suzuki, Lee \& McPhail, 1982), cantoniensistriol triacetate (Mak, Chang \& Chang, 1982) and in $[3 \beta(E)]$-acetoxyolean12 -en- $28 \beta$-oic acid etc. (Roques, Declercq \& Germain, 1978). The lengthening of the $\mathrm{C} 8-\mathrm{C} 14$ $[1.604$ (4) $\AA], \mathrm{C} 9-\mathrm{C} 10[1.581(4) \AA]$ and $\mathrm{C} 4-\mathrm{C} 5$ [ 1.572 (4) $\AA$ ] bonds is in accord with expectations and is due to steric crowding around C8, C9, C14

[^0]Table 2. Bond distances $(\AA)$ and angles $\left(^{\circ}\right)$ with e.s.d.'s in parentheses

| $\mathrm{Ol}-\mathrm{C} 22$ | 1.459 (4) | $\mathrm{Cl1}-\mathrm{Cl2}$ | 1.502 (5) |
| :---: | :---: | :---: | :---: |
| $\mathrm{Ol}-\mathrm{C} 31$ | 1.337 (4) | $\mathrm{Cl2-Cl3}$ | 1.334 (5) |
| O2-C3 | $1 \cdot 221$ (4) | Cl3-Cl4 | 1.529 (4) |
| O3-C28 | 1.327 (4) | C13-C18 | 1.534 (4) |
| O4-C28 | $1 \cdot 199$ (4) | C14-C15 | 1.551 (5) |
| O5-C31 | 1.202 (5) | C14-C27 | 1.543 (5) |
| $\mathrm{Cl}-\mathrm{C} 2$ | 1.506 (5) | $\mathrm{Cl5}-\mathrm{Cl} 6$ | 1.543 (4) |
| $\mathrm{Cl}-\mathrm{Cl} 10$ | 1.536 (5) | C16-C17 | 1.543 (4) |
| $\mathrm{C} 2-\mathrm{C} 3$ | 1.459 (5) | C17-C18 | 1.552 (4) |
| C3-C4 | 1.515 (5) | C17-C22 | 1.547 (4) |
| C4-C5 | 1.572 (4) | C17-C28 | 1.528 (5) |
| C5-C6 | 1.526 (5) | C18--C19 | 1.541 (5) |
| C5-C10 | 1.545 (5) | $\mathrm{C} 19-\mathrm{C} 20$ | 1.538 (4) |
| C6-C7 | 1.509 (4) | C20-C21 | 1.532 (5) |
| C7-C8 | 1.542 (4) | C20-C29 | 1.543 (6) |
| C8-C9 | 1.548 (4) | $\mathrm{C} 20-\mathrm{C} 30$ | 1.547 (5) |
| C8-C14 | 1.601 (4) | C21-C22 | 1.525 (5) |
| C8-C26 | 1.546 (5) | C31-C32 | 1.508 (6) |
| C9-C10 | 1.581 (4) | C32-C33 | $1 \cdot 338$ (7) |
| C9-Cll | 1.540 (5) | C32-C35 | 1.519 (8) |
| $\mathrm{C} 10-\mathrm{C} 25$ | 1.532 (5) | C33-C34 | $1 \cdot 40$ (1) |
| C22-O1-C31 | 117.9 (3) | C13-C14-C15 | $110 \cdot 1$ (2) |
| $\mathrm{C} 2-\mathrm{Cl}-\mathrm{Cl} 0$ | 115.4 (3) | C13-C14-C27 | $109 \cdot 1$ (3) |
| $\mathrm{Cl}-\mathrm{C} 2-\mathrm{C} 3$ | $117 \cdot 6$ (3) | C15-C14-C27 | 106.7 (3) |
| O2-C3-C2 | 119.3 (3) | C14-C15-C16 | 113.4 (3) |
| $\mathrm{O} 2-\mathrm{C} 3-\mathrm{C} 4$ | 119.9 (3) | C15-C16-C17 | $111 \cdot 1$ (2) |
| C2-C3-C4 | 120.9 (3) | C16-C17-C18 | $109 \cdot 2$ (2) |
| C3-C4-C5 | 110.8 (3) | $\mathrm{C} 16-\mathrm{C} 17-\mathrm{C} 22$ | 109.8 (2) |
| C4-C5-C6 | 112.8 (3) | C16-C17-C28 | 109.0 (2) |
| C4-C5-C10 | 114.6 (3) | C18-C17-C22 | 112.0 (2) |
| C6-C5-C10 | 111.6 (3) | C18-C17-C28 | $110 \cdot 5$ (2) |
| $\mathrm{C5}-\mathrm{C} 6-\mathrm{C} 7$ | 110.9 (3) | C22-C17-C28 | $106 \cdot 3$ (2) |
| C6- $\mathrm{C} 7-\mathrm{C} 8$ | $113 \cdot 3$ (3) | C13-C18-C17 | 111.6 (2) |
| C7-C8-C9 | $110 \cdot 3$ (2) | C13-C18-C19 | 111.1 (3) |
| C7--C8-C14 | 109.9 (2) | C17-C18-C19 | 111.9 (2) |
| C7-C8-C26 | 108.1*(3) | C18-C19-C20 | 115.7 (3) |
| $\mathrm{C} 9-\mathrm{C} 8-\mathrm{Cl} 4$ | 107.4 (2) | $\mathrm{C} 19-\mathrm{C} 20-\mathrm{C} 21$ | 108.4 (3) |
| C9-C8-C26 | 111.7 (3) | C19-C20-C29 | 1119 (3) |
| C14-C8-C26 | 109.5 (2) | $\mathrm{C} 19-\mathrm{C} 20-\mathrm{C} 30$ | 109.1 (3) |
| $\mathrm{C} 8-\mathrm{C} 9-\mathrm{Cl} 0$ | 118.0 (3) | C21-C20-C29 | 112.5 (3) |
| C8-C9-Cl1 | 109.1 (3) | C21-C20-C30 | 107.8 (3) |
| $\mathrm{C} 10-\mathrm{C}-\mathrm{Cll}$ | 111.9 (3) | C29-C20-C30 | 107.0 (3) |
| $\mathrm{Cl}-\mathrm{Cl} 0-\mathrm{C} 5$ | $107 \cdot 9$ (3) | C20-C21-C22 | $115 \cdot 3$ (3) |
| $\mathrm{Cl}-\mathrm{Cl} 10-\mathrm{C} 9$ | 107.1 (3) | $\mathrm{Ol}-\mathrm{C} 22-\mathrm{C} 17$ | $105 \cdot 6$ (2) |
| $\mathrm{Cl}-\mathrm{C} 10-\mathrm{C} 25$ | $107 \cdot 0$ (3) | $\mathrm{O} 1-\mathrm{C} 22-\mathrm{C} 21$ | 109.9 (2) |
| $\mathrm{C5}-\mathrm{Cl0}-\mathrm{C} 9$ | $108 \cdot 1{ }^{\prime}(3)$ | C17-C22-- 21 | 114.0 (3) |
| $\mathrm{C} 5-\mathrm{C} 10-\mathrm{C} 25$ | 112.1 (3) | O3-C28-O4 | $123 \cdot 1$ (3) |
| $\mathrm{C} 9-\mathrm{C} 10-\mathrm{C} 25$ | $114 \cdot 2$ (3) | O3-C28-C17 | 111.7 (3) |
| C9-C11-C12 | 111.5 (3) | O4-C28-C17 | $125 \cdot 2$ (3) |
| C11-C12-C13 | 126.4 (3) | $\mathrm{Ol}-\mathrm{C} 31-\mathrm{O} 5$ | $122 \cdot 6$ (4) |
| C12-C13-C14 | $121 \cdot 3$ (3) | $\mathrm{Ol}-\mathrm{C} 31-\mathrm{C} 32$ | 109.8 (3) |
| C12-C13-C18 | 117.4 (3) | $\mathrm{O}-\mathrm{C} 31-\mathrm{C} 32$ | 127.6 (3) |
| C14-C13-C18 | $121 \cdot 3$ (3) | C31-C32-C33 | 119.6 (4) |
| C8-C14--C13 | 109.1 (2) | C31-C32-C35 | 116.5 (4) |
| C8-C14-Cl5 | 110.0 (2) | C33-C32-C35 | 123.7 (5) |
| C8-C14-- ${ }^{-} 27$ | 111.7(2) | C32-C33-C34 | $130 \cdot 8$ (7) |

and C 23 , while $\mathrm{C} 11-\mathrm{C} 12$ [1-502 (5) $\AA$ ] and $\mathrm{C} 6-\mathrm{C} 7$ [ $1.509(4) \AA$ ] appear to be shortened. Fig. 1 shows a view of the molecule.

Three of the five six-membered rings $(B, D$ and $E)$ are in the chair conformation, while rings $A$ and $C$ are in sofa and half-chair conformations, respectively (Duax, Weeks \& Rohrer, 1976) (Fig. 1). The presence of a double bond in ring $C$ accounts for its twist conformation. Furthermore, it has been reported that the keto functionality will generally result in ring $A$ adopting a twisted boat conformation (Corbett, Simpson, Goh, Nicholson, Wilkins \& Robinson, 1982). The $A / B$ and $B / C$ rings are trans fused while the $D / E$ rings are cis fused as in the case of methylmicromerol bromoacetate (Stout \& Stevens, 1963) and cantoniensistriol triacetate (Mak, Chang \& Chang, 1982).


Fig. 1. Stereoview of the molecule.
Substitution at the 22 position is $\beta$-axial. The absolute configuration of the structure has not been established but is given as in the related structure. The packing of the molecule is stabilized by $\mathrm{O} 3-\mathrm{H} \cdots \mathrm{O} 2[2.704(4) \AA]$ type hydrogen bonds. The $\mathrm{C} 28-\mathrm{O} 3 \cdots \mathrm{O} 2$ angle is $115 \cdot 0(3)^{\circ}$.

## References

Corbett, R. E., Simpson, J., Goh, E. M., Nicholson, B. K., Wilkins, A. L. \& Robinson, W. T. (1982). J. Chem. Soc. Perkin Trans. 2, pp. 1339-1343.
Duax, W. L., Weeks, C. M. \& Rohrer, D. C. (1976). Topics in Stereochemistry, edited by N. L. Allinger \& E. L. Elliel, ch. 9, pp. 271-383. New York: John Wiley.
Frenz, B. A. (1978). The Enraf-Nonius CAD4 SDP - A RealTime System for Concurrent X-ray Data Collection and Crystal Structure Solution. In Computing in Crystallography, edited by H. Schenk, R. Olthof-Hazekamp, H. van Koningsveld \& G. C. Bassi. Delft Univ. Press.

Mak, T. C. W., Chang, T.-C. \& Chang, H.-M. (1982). J. Chem. Soc. Chem. Commun. pp. 785-786.
Nozaki, H., Suzuki, H., Lee, K. H. \& McPhail, A. T. (1982). J. Chem. Soc. Chem. Commun. pp. 1048-1051.
Ravin, L. J. (1985). Preformulations. In Remington's Pharmaceutical Sciences, 17th ed., edited by A. R. Gennaro. Easton, PA: Mack.
Roques, P. R., DeclercQ, J.-P. \& Germain, G. (1978). Acta Cryst. B34, 2367-2370.
Sharma, O. P., Dawra, P. K. \& Makkar, H. P. S. (1987). Toxicol. Lett. 37, 165-172.
Sharma, O. P., Dawra, R. K. \& Makkar, H. P. S. (1988). Toxicol. Lett. 42, 29-37.
Sharma, O. P., Makkar, H. P. S. \& Dawra, R. K. (1988). Toxicon, 26, 975-987.
Stout, G. H. \& Stevens, K. L. (1963). J. Org. Chem. 28, 1259-1262.


[^0]:    * Lists of structure factors, anisotropic thermal parameters, H -atom parameters, torsion angles and least-squares-planes data have been deposited with the British Library Document Supply Centre as Supplementary Publication No. SUP 53452 ( 28 pp.). Copies may be obtained through The Technical Editor, International Union of Crystallography, 5 Abbey Square, Chester CHI 2HU, England.

